

**Structural Elucidation of a Novel Erythromycin,
13-Cyclopentyl-13-desethyl-erythromycin B,
from a Recombinant *Saccharopolyspora
erythraea* Strain, NRRL 2338 pIG/1**

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In a preceding paper,¹⁾ we described the production and isolation of 13-cyclopentyl-13-desethyl-erythromycin B, **1**, from the genetically modified organism *Saccharopolyspora erythraea* NRRL 2338 pIG/1. This paper describes the NMR studies used to confirm the structure of **1**, and to establish the relative stereochemistry of the substituent at the C-13 position.

The structure of **1** was confirmed by analysis of NMR data, obtained at 500 MHz in CDCl₃. ¹H-¹H coupling systems within the molecule were revealed by TOCSY spectra, obtained with 30 ms and 80 ms mixing times. Direct (¹J) and long-range (²J and ³J) ¹H-¹³C correlations were revealed by gradient-selected edited HSQC and gradient-enhanced HMBC experiments respectively. Key HMBC correlations, which enabled the presence of the macrolide ring in **1** to be confirmed, are shown in Figure 1.

The positions of attachment of the cladinose and desosamine sugars were confirmed by virtue of HMBC

correlations from H-1' to C-5 and from H-1'' to C-3. The chemical shifts of C-1' and C-1'' identified these two carbons as anomeric. The ¹H-¹H coupling systems around these two sugars and the rest of the molecule were identified by TOCSY experiments.

A high resolution electrospray mass spectrum obtained for **1** revealed a sodiated molecular ion with formula [C₄₀H₇₁NO₁₂Na]⁺. This corresponds to a molecular formula of C₄₀H₇₁NO₁₂, which is consistent with the identity of **1** as 13-cyclopentyl-13-desethyl-erythromycin B.

A double-quantum COSY experiment showed that H-13 was coupled to the H-14 signal at 2.15 ppm, and therefore confirmed the linkage of C-13 to C-14. H-14 was then identified as a methine hydrogen, from the edited HSQC experiment. A 1D TOCSY experiment showed that the H-15~H-18 hydrogens also belonged to the coupling system which included H-13 and H-14, and the edited HSQC experiment confirmed that the four carbons C-15, 16, 17 and 18 were all methylenes. Attachment of these four methylenes *via* C-15 to C-14 accounted for all the atoms of the molecular formula of C₄₀H₇₁NO₁₂, and cyclisation of the C-18 methylene onto C-14 accounted for all the unfulfilled valencies of the molecule. The structure of **1** was therefore confirmed as 13-cyclopentyl-13-desethyl-erythromycin B. Full ¹H and ¹³C NMR assignments for **1** are listed in Table 1. These are comparable with those reported in the literature²⁾ for erythromycin A.

The conformation and relative stereochemistry of **1** was investigated as follows. Erythromycin analogues crystallise in one of two preferred conformations,^{3,4)} which are also present in the solution state. The preferred solution conformation of a given derivative can be

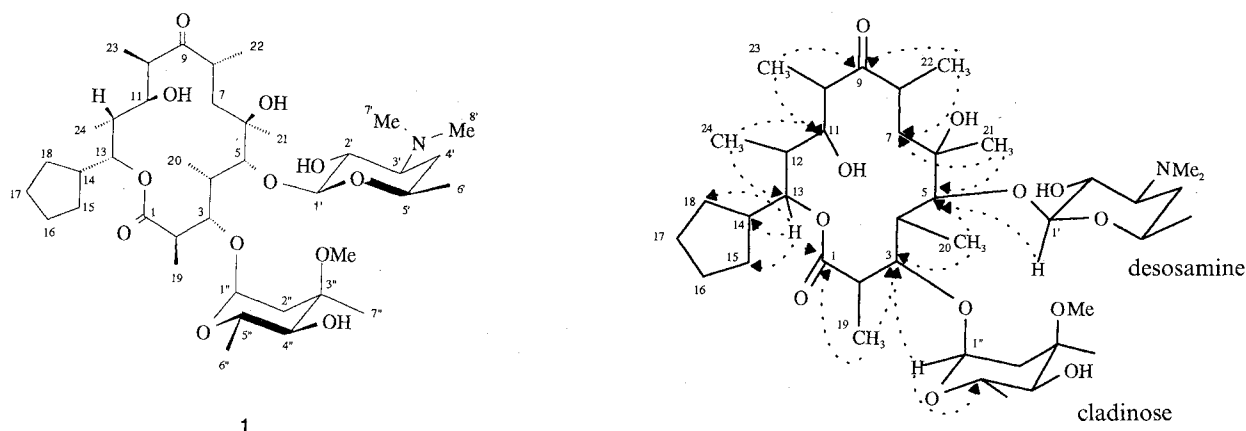
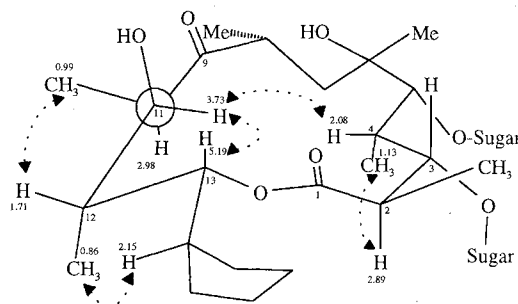


Fig. 1. Key HMBC correlations of **1**.

Table 1. ^1H and ^{13}C assignments for **1**.

Atom number	^{13}C chemical shift, from ^{13}C NMR spectrum	^1H chemical shift and multiplicity
1	176.0	—
2	45.0	2.89 1H dq $J=9.4, 7.1$
3	80.4	4.02 1H dd $J=9.4, 1.7$
4	39.2	2.08 1H multiplet
5	83.8	3.59 1H d $J=7.4$
6	75.4	—
7	38.0	2.00 1H dd $J=14.7, 10.8$ <i>ca.</i> 1.65 1H multiplet
8	45.0	2.71 1H dqd $J=10.6, 6.8, 2.6$
9	<i>ca.</i> 220.0	—
10	38.9 ^a	2.98 1H qd $J=6.8, 1.5$
11	69.4	3.73 1H dd $J=9.9, 1.2$
12	38.8 ^a	1.71 1H multiplet
13	78.3	5.19 1H dd $J=10.5, 1.0$
14	41.7	2.15 1H br sextet
15	30.4	<i>ca.</i> 1.69 1H multiplet <i>ca.</i> 1.21 1H multiplet
16	25.4	<i>ca.</i> 1.63 1H multiplet <i>ca.</i> 1.52 1H multiplet
17	25.1	<i>ca.</i> 1.63 1H multiplet <i>ca.</i> 1.52 1H multiplet
18	29.0	<i>ca.</i> 1.69 1H multiplet <i>ca.</i> 1.21 1H multiplet
19	15.8	1.18 3H d $J=7.1$
20	9.24	1.13 3H d $J=7.0$
21	27.4	1.46 3H s
22	18.5	1.14 3H d $J=6.8$
23	9.5	0.99 3H d $J=6.8$
24	9.16	0.86 3H d $J=7.1$
1'	103.2	4.43 1H d $J=7.3$
2'	70.9	3.24 1H dd $J=10.3, 7.3$
3'	65.4	2.51 1H ddd $J=12.0, 10.6, 4.1$
4'	29.0	<i>ca.</i> 1.68 1H multiplet <i>ca.</i> 1.24 1H multiplet
5'	68.8	3.50 1H br sextet
6'	21.5	1.22 3H d $J=6.1$
7',8'	40.1	2.32 2 × 3H s
1''	96.5	4.90 1H d $J=4.6$
2''	35.1	2.36 1H d $J=15.2$ + small (< 1 Hz) 1.58 1H multiplet
3''	72.6	—
4''	78.0	3.02 1H d $J=9.1$
5''	65.6	4.01 1H multiplet
6''	18.7	1.29 3H d $J=6.3$
7''	21.4	1.24 3H s
8''	49.5	3.31 3H s

^a Assignments for signals with asterisks may be interchangeable.

Fig. 2. Approximate representation of conformation of **1**, showing how observed ROESY correlations are consistent with this conformation.

recognised by key NOE interactions,^{5,6)} and by analysis of ^1H - ^1H chemical shifts and coupling constants.⁷⁾ Thus, in **1**, a ROESY correlation between H-4 and H-11 indicated that **1** adopted a similar molecular conformation to the 'folded out' conformation revealed by X-ray crystallography of erythromycin A hydroiodide dihydrate³⁾ (Figure 2). No evidence for the presence of the alternative erythromycin conformation⁴⁾ (e.g. an H-11-H-3 correlation) was seen in the ROESY spectrum. All other observed ROESY correlations and ^1H - ^1H couplings were consistent with **1** existing in a similar conformation to that of erythromycin A hydroiodide dihydrate.³⁾ A ROESY correlation between H-11 and H-13 enabled the relative stereochemistry at C-13 to be identified as the same as that in the 13-ethylerythromycins, as shown Figure 2.

The ROESY and ^1H - ^1H coupling data therefore indicate that the relative stereochemistry of **1** is as shown above.

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